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# Molecular hydrogen as a preventive and therapeutic medical gas: initiation, development and potential of hydrogen medicine



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## ABSTRACT

Molecular hydrogen ( $H_2$ ) has been accepted to be an inert and nonfunctional molecule in our body. We have turned this concept by demonstrating that  $H_2$  reacts with strong oxidants such as hydroxyl radical in cells, and proposed its potential for preventive and therapeutic applications.  $H_2$  has a number of advantages exhibiting extensive effects:  $H_2$  rapidly diffuses into tissues and cells, and it is mild enough neither to disturb metabolic redox reactions nor to affect signaling reactive oxygen species; therefore, there should be no or little adverse effects of  $H_2$ . There are several methods to ingest or consume  $H_2$ ; inhaling  $H_2$  gas, drinking  $H_2$ -dissolved water ( $H_2$ -water), injecting  $H_2$ -dissolved saline ( $H_2$ -saline), taking an  $H_2$  bath, or dropping  $H_2$ -saline into the eyes. The numerous publications on its biological and medical benefits revealed that  $H_2$  reduces oxidative stress not only by direct reactions with strong oxidants, but also indirectly by regulating various gene expressions. Moreover, by regulating the gene expressions,  $H_2$  functions as an anti-inflammatory and anti-apoptotic, and stimulates energy metabolism. In addition to growing evidence obtained by model animal experiments, extensive clinical examinations were performed or are under investigation. Since most drugs specifically act to their targets,  $H_2$  seems to differ from conventional pharmaceutical drugs. Owing to its great efficacy and lack of adverse effects,  $H_2$  has promising potential for clinical use against many diseases.

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## Contents

|   |   |
|---|---|
| 1. Introduction . . . . .   | 1 |
| 2. Oxidative stress as pathogenic sources and physiological roles . . . . .                 | 2 |
| 3. Discovery of the biological effects of molecular hydrogen . . . . .                      | 3 |
| 4. Methods of how to ingest molecular hydrogen . . . . .                                    | 3 |
| 5. Monitoring the movement of molecular hydrogen . . . . .                                  | 4 |
| 6. Effects of molecular hydrogen and clinical examinations . . . . .                        | 4 |
| 7. Possible molecular mechanisms underlying various effects of molecular hydrogen . . . . . | 6 |
| 8. Comparison with the other medical gasses . . . . .                                       | 8 |
| 9. Concluding remarks . . . . .   | 8 |
| Conflict of Interest Statement . . . . .  | 8 |
| References . . . . .  | 9 |

## 1. Introduction

Molecular hydrogen ( $H_2$ , dihydrogen, or hydrogen gas) has been accepted to behave as an inert gas at body temperature in mammalian cells. In fact,  $H_2$  seems to react with no biological compound, including oxygen gas in the absence of catalysts at body temperature. On the

other hand, in some bacteria,  $H_2$  is enzymatically catabolyzed as an energy source for providing electrons, or is a product of some types of anaerobic metabolism. These reactions are usually catalyzed by iron- or nickel-containing enzymes called hydrogenases. In contrast, mammals have no functional hydrogenase genes (Fritsch et al., 2013). Thus, it has been believed that  $H_2$  is nonfunctional in our cells.

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We turned this concept in a publication in 2007 that H<sub>2</sub> acts as a therapeutic and preventive antioxidant by selectively reducing highly strong oxidants such as hydroxyl radical ( $\bullet\text{OH}$ ) and peroxynitrite ( $\text{ONOO}^-$ ) in cells, and that H<sub>2</sub> exhibits cytoprotective effects against oxidative stress (Ohsawa et al., 2007). Since then, a large number of studies have explored therapeutic and preventive effects of H<sub>2</sub>. These published papers cover many biological effects against oxidative stress in almost all organs (Ohta, 2011, 2012). Moreover, it has been revealed that H<sub>2</sub> has more functions, including anti-inflammatory, anti-apoptotic, and anti-allergic effects, and that H<sub>2</sub> stimulates energy metabolism, in most tissues of model animals. Until 2013, the number of publications on its biologically or medically beneficial effects had been increasing and had surpassed ~300 as shown in Fig. 1. Previous review articles mainly introduced various cellular and animal experiments (Ohta, 2011, 2012).

In addition to publications on model animal experiments, more than 10 papers on clinical examinations have been published. Thus, this article will review the results of recent clinical examinations toward actual applications. Moreover, this review article will look back the discovery process of the biological effects of H<sub>2</sub>, and propose the possible mechanisms to explain the effects of H<sub>2</sub>.

## 2. Oxidative stress as pathogenic sources and physiological roles

### 2.1. Oxidative stress as pathogenic sources

Reactive oxygen species (ROS) are generated inside the body throughout our daily lives as a side-product of energy metabolism by oxidative phosphorylation in every aerobic organism. Occasionally, excess ROS are produced, such as smoking or air pollution, exposure to ultraviolet or irradiation rays, hard exercise, physical or psychological stress, and so on (Liu et al., 1996; Agarwal, 2005; Harma et al., 2006; Tanriverdi et al., 2006; Grassi et al., 2010). When ROS are produced excessively or endogenous antioxidant capacity is diminished, indiscriminate oxidation elicits harmful effects, resulting in “oxidative stress”. Acute oxidative stress arises from various different situations: inflammation, ischemia reperfusion in cardiac or cerebral infarction, organ transplantation, and cessation of operative bleeding, or others (Ferrari et al., 1991; Vaziri & Rodriguez-Iturbe, 2006; Reuter et al., 2010). Mounting evidence has established strong links between chronic oxidative stress and a wide variety of pathologies, including malignant diseases, diabetes mellitus, atherosclerosis, and chronic inflammatory processes as well as many neurodegenerative diseases and the aging process (Andersen, 2004; Bagul & Banerjee, 2013; El Assar et al., 2013; Kim & Byzova, 2014). Under normal conditions, ROS induced by strenuous exercise results in muscle fatigue (Westerblad & Allen, 2011).

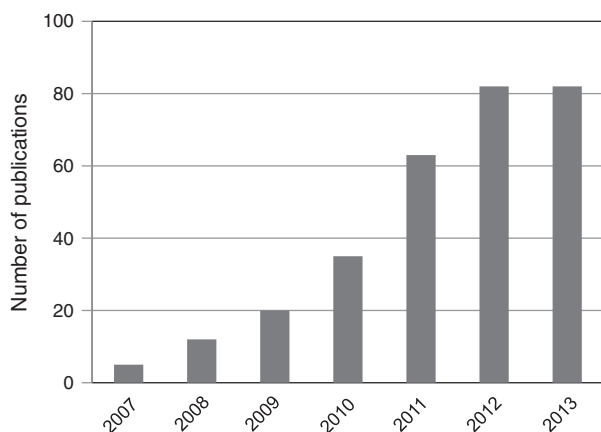


Fig. 1. The number of publications on biological effects of molecular hydrogen in each year.

### 2.2. The process of generation of oxygen species

As a first step in generating ROS, superoxide anion radicals ( $\text{O}_2^{\bullet-}$ ) are the primary ROS mostly generated by electron leakage from the mitochondrial electron transport chain (Finkel & Holbrook, 2000; Turrens, 2003; Andersen, 2004; Lin & Beal, 2006). Superoxide dismutase (SOD) enzymatically converts  $\text{O}_2^{\bullet-}$  to hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), which is metabolized to generate water ( $\text{H}_2\text{O}$ ). Very reactive  $\bullet\text{OH}$  is generated from  $\text{H}_2\text{O}_2$  via the Fenton or Weiss reaction in the presence of catalytically active metals, such as  $\text{Fe}^{2+}$  and  $\text{Cu}^+$  (Halliwell & Gutteridge, 1992). Reaction of  $\text{O}_2^{\bullet-}$  with nitric oxide ( $\bullet\text{NO}$ ) generates  $\text{ONOO}^-$ , which is a very active nitrogen species (RNS) (Radi, 2013).  $\bullet\text{OH}$  is the major cause of the oxidation and destruction of biomolecules by the direct reaction or by triggering the chain reaction of free radicals (Lipinski, 2011). Ionizing radiation, including cosmic rays also generates  $\bullet\text{OH}$  as a damaging intermediate through interaction with water, a process termed radiolysis (Schoenfeld et al., 2011, 2012).

Other enzymes, including NADPH oxidases, cytochrome p450s, lipoxygenase, cyclooxygenase, and xanthine oxidase, also participate in ROS generation in the immuno- or detoxifying-system (Droge, 2002).

### 2.3. Physiological roles of $\text{H}_2\text{O}_2$

As mentioned above, ROS had historically been believed to cause cellular damage and to lack physiological functions. Indeed, accumulation of oxidative damage by ROS has been linked to multiple pathologies, as mentioned above. Cellular redox homeostasis is, however, a delicate balance between ROS production and the antioxidant system (Bashan et al., 2009; Brewer et al., 2013). Oxidative stress is now appreciated to function as signaling molecules to regulate a wide variety of physiology (Liu et al., 2005; Bell et al., 2007a,b).  $\text{H}_2\text{O}_2$  was shown to be required for cytokine, insulin, growth factor, AP-1, c-Jun N-terminal kinase 1 (JNK1), p53, and nuclear factor kappa B (NF- $\kappa$ B) signaling and to promote phosphatase-inactivation by cysteine oxidation (Finkel, 1998; Chandel et al., 2000a,b). These reactions provide a plausible biochemical mechanism by which ROS can impinge on signaling pathways. There have been numerous reports highlighting the importance of ROS-dependent signaling in a variety of systems (Salganik, 2001; Sauer et al., 2001; Collins et al., 2012).

Additionally, oxidative stress caused by  $\text{H}_2\text{O}_2$  and  $\bullet\text{NO}$  induces enzymes involved in anti-oxidation and tolerance to protect cells against oxidative stress (Endo et al., 2009; Ristow & Zarse, 2010). For example, translocation of NF-E2-related factor 2 (Nrf2) into the nucleus leads to the regulation of gene expression involved in defense systems against oxidative stress (Jazwa & Cuadrado, 2010) and other toxic sources including heavy metals (Gan & Johnson, 2013). Moreover,  $\text{H}_2\text{O}_2$  is a key factor to regulate cellular differentiation (Tsukagoshi et al., 2010; Tormos et al., 2011), the immune system (West et al., 2011; Zhou et al., 2011), autophagy (Li et al., 2012; Garg et al., 2013) and apoptosis (Mates et al., 2012). Thus, it is crucial not to completely eliminate  $\text{H}_2\text{O}_2$  to maintain homeostasis.

### 2.4. Exploration of an ideal antioxidant

Although antioxidant therapy or prevention of various diseases is expected owing to the clinical importance of oxidative damage, antioxidants have been of limited therapeutic success (Steinhubl, 2008). Antioxidant supplements have exhibited little effect on preventing cancer, myocardial infarction and atherosclerosis, but rather conversely have increased mortality (Bjelakovic et al., 2007; Hackam, 2007; Brambilla et al., 2008; Steinhubl, 2008; Hercberg et al., 2010); thus, it is very important to be aware of side effects in developing an effective antioxidant for the prevention of oxidative stress-related diseases.

Under these situations, an ideal antioxidant molecule is expected to mitigate excess oxidative stress, but not disturb the redox homeostasis. In other words, the ideal molecule should not reduce signaling molecules

such as  $\text{H}_2\text{O}_2$  but should effectively reduce strong oxidants such as  $\bullet\text{OH}$ . After experiments, we have reached the current conclusion that the ideal antioxidant could be  $\text{H}_2$ .

### 3. Discovery of the biological effects of molecular hydrogen

During the process of searching for the ideal antioxidant, we infused  $\text{H}_2$  into culture medium without changing pH,  $\text{O}_2$  and  $\text{CO}_2$  concentrations or other conditions. When cultured PC12 cells were exposed to oxidative stress by treatment with antimycin A, an inhibitor of the mitochondrial electron translocation chain, the cells apparently shrank and extended short fibers in response to oxidative stress (Fig. 2A). In contrast, when the cells were treated with the inhibitor in the presence of  $\text{H}_2$ , the cells did not change their shape (Fig. 2B). In  $\text{H}_2$ -degassed medium from  $\text{H}_2$ -medium, the cells again responded to oxidative stress. This finding indicates that  $\text{H}_2$  affected no components in the original medium, but directly acted to the cells. By this finding of the first experiment, we foresaw that  $\text{H}_2$  has great potential for actual clinical use.

After this experiment, we tried to identify the target of  $\text{H}_2$  in cultured cells.  $\text{H}_2$  dissolved in culture medium did not change the cellular levels of  $\text{O}_2\bullet^-$  and  $\text{H}_2\text{O}_2$ , as judged by the fluorescent signals of MitoSOX and dichlorofluorescein-diacetate (DCF-DA), respectively. Additionally,  $\text{H}_2$  did not decrease the cellular level of  $\bullet\text{NO}$ . In contrast,  $\text{H}_2$  treatment significantly decreased levels of  $\bullet\text{OH}$ , as judged by the decrease in the fluorescent signal of hydroxyphenyl fluorescein (HPF) (Setsukinai et al., 2003). Moreover, the decrease in the cellular  $\bullet\text{OH}$  level by  $\text{H}_2$  was confirmed by spin trapping technology (Halliwell & Gutteridge, 1992).

The selective reduction of ROS can be explained by the marked oxidative strength of  $\bullet\text{OH}$ , as shown in Fig. 3, which was profiled according to published data (Setsukinai et al., 2003). This means that  $\bullet\text{OH}$  is strong enough to react with even inert  $\text{H}_2$ , but that  $\text{O}_2\bullet^-$ ,  $\text{H}_2\text{O}_2$ , and  $\bullet\text{NO}$  are insufficient to react with  $\text{H}_2$  according to their activities. In other words,  $\text{H}_2$  is mild enough neither to disturb metabolic redox reactions nor to affect ROS that function in cellular signaling.

## 4. Methods of how to ingest molecular hydrogen

### 4.1. Safety of molecular hydrogen

Previous review articles have summarized the nature of  $\text{H}_2$  as a chemical element (Ohta, 2011, 2012). Here, the following point should be emphasized regarding the safety of  $\text{H}_2$  for medical use:  $\text{H}_2$  gas is flammable only at temperatures higher than  $527^\circ\text{C}$ , and explodes by a rapid chain reaction with  $\text{O}_2$  only in the explosive range of  $\text{H}_2$  concentration (4–75%, vol/vol). Thus,  $\text{H}_2$  can be used for medical applications without surplus worries by several ingestion methods because

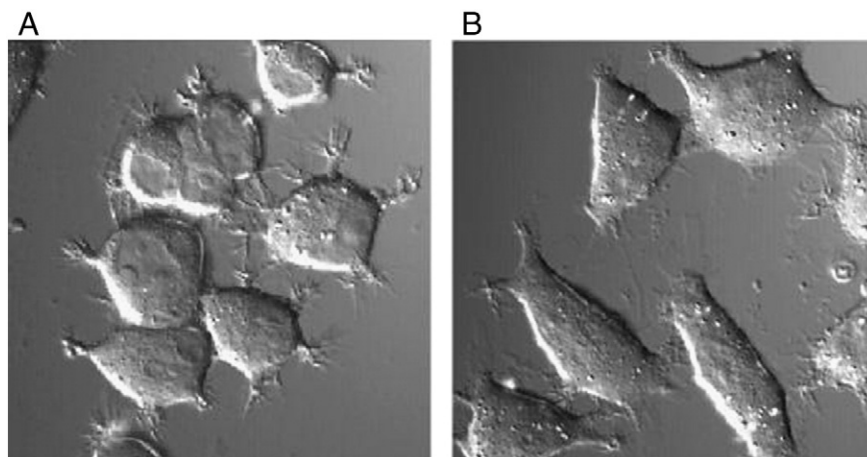


Fig. 2. Photographs of cultured PC12 exposed to oxidative stress by treatment with antimycin A without (A) or with (B) hydrogen in media.

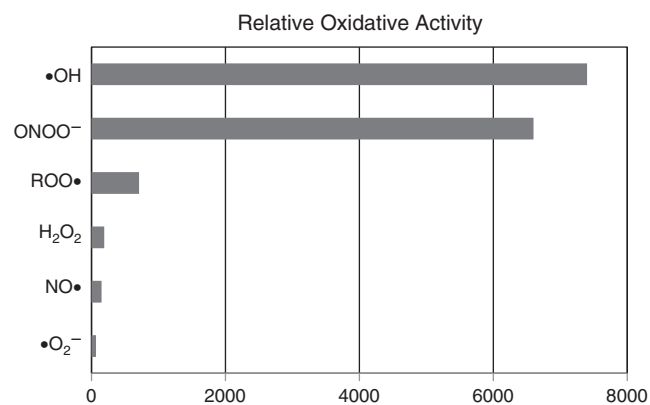


Fig. 3. Relative oxidative activities in each reactive oxygen and nitrogen species. This graph is illustrated based on data from a previous publication (Setsukinai et al., 2003).

inhalation of 1–4%  $\text{H}_2$  gas exhibits great efficacy (Ohsawa et al., 2007; Hayashida et al., 2008).

### 4.2. Inhalation of hydrogen gas

Inhalation of  $\text{H}_2$  gas is a straightforward therapeutic method.  $\text{H}_2$  gas can be inhaled through a ventilator circuit, facemask or nasal cannula. Since inhaled  $\text{H}_2$  gas acts rapidly, it may be suitable for defense against acute oxidative stress. In particular, inhalation of gas does not affect blood pressure (Ohsawa et al., 2007); on the other hand, drip infusion of drugs increases blood pressure and causes serious obstacles during the treatment of myocardial infarction.

By a clinical examination, Ono et al. showed that inhalation of 3–4%  $\text{H}_2$  gas reached a plateau at approximately  $10\text{--}20\ \mu\text{M}$  in the arterial and venous blood, respectively, in about 20 min and affected no physiological parameters, suggesting no adverse effects (Ono et al., 2012a).

### 4.3. Oral ingestion by drinking hydrogen water

Inhalation of  $\text{H}_2$  gas may be unsuitable or impractical for continuous  $\text{H}_2$  consumption in daily life for preventive use. In contrast, solubilized  $\text{H}_2$  ( $\text{H}_2$ -dissolved water; namely,  $\text{H}_2$ -water) may be beneficial since it is a portable, easily administered and safe way to ingest  $\text{H}_2$  (Nagata et al., 2009; Nakashima-Kamimura et al., 2009).  $\text{H}_2$  can be dissolved in water up to  $0.8\ \text{mM}$  ( $1.6\ \text{mg/L}$ ) under atmospheric pressure at room temperature without changing pH. Although *ad libitum* drinking saturated  $\text{H}_2$ -water was more effective than diluted one,  $80\ \mu\text{M}$  of  $\text{H}_2$ -water was still effective for improving obesity (Kamimura et al., 2011).

H<sub>2</sub>-water can be made by several methods: infusing H<sub>2</sub> gas into water under high pressure, electrolyzing water to producing H<sub>2</sub>, and reacting of magnesium metal or its hydride with water. These methods may be applicable not only to water but also to other solvents. H<sub>2</sub> penetrates the glass and plastic walls of any vessels in a short time, while aluminum containers are able to retain hydrogen gas for a long time.

#### 4.4. Injection of hydrogen-saline

H<sub>2</sub> is intravenously or intraperitoneally injectable as H<sub>2</sub>-saline (H<sub>2</sub>-dissolved saline), which allows the delivery of H<sub>2</sub> with great efficacy in model animals (Cai et al., 2009; Sun et al., 2011a,b; G.M. Li et al., 2013).

Nagatani et al. performed an open-label, prospective, non-randomized study of intravenous H<sub>2</sub> administration in 38 patients hospitalized for acute ischemic stroke. All patients received an H<sub>2</sub> intravenous solution (200 mL twice a day) immediately after the diagnosis of acute ischemic stroke. Data from this study indicated that an H<sub>2</sub> intravenous solution is safe for patients with acute cerebral infarction, including patients treated with tissue-plasminogen activator (t-PA) (Nagatani et al., 2013).

#### 4.5. Direct incorporation of molecular hydrogen by diffusion: Eye-drop, bath and cosmetics

Alternatively, H<sub>2</sub>-loaded eye drops were prepared by dissolving H<sub>2</sub> in saline and directly administering to the ocular surface (Oharazawa et al., 2010; Kubota et al., 2011).

H<sub>2</sub> delivery to cardiac grafts during cold preservation using a hydrogen-supplemented water bath efficiently ameliorated myocardial injury due to cold ischemia and reperfusion. This device to saturate organs with H<sub>2</sub> during cold storage merits further investigation for possible therapeutic and preventative use during transplantation (Noda et al., 2013).

H<sub>2</sub> should easily penetrate the skin and is distributed throughout the body via blood flow. Thus, taking a warm water bath with dissolved H<sub>2</sub> is a method of incorporating H<sub>2</sub> into the body in daily life. Indeed, powders that produce H<sub>2</sub> baths are commercially available in Japan.

### 5. Monitoring the movement of molecular hydrogen

H<sub>2</sub> has a number of advantages as a potential antioxidant. It has favorable distributing characteristics with its own physical ability to penetrate biomembranes and diffuse into the cytosol. H<sub>2</sub> rapidly reaches the nucleus and mitochondria to protect nuclear DNA and mitochondria (Ohsawa et al., 2007). Moreover, H<sub>2</sub> passes through the blood-brain barrier although most antioxidant compounds cannot.

H<sub>2</sub> in blood can be monitored by the following method; venous or arterial blood is collected in a closed aluminum bag with no dead space, followed by the addition of a definite volume of air into the bag. H<sub>2</sub> in the air phase transferred from the blood can be measured by gas chromatography. The inhalation of H<sub>2</sub> actually increased H<sub>2</sub> dissolved in arterial blood in a H<sub>2</sub> gas concentration-dependent manner, and the H<sub>2</sub> levels in venous blood were lower than in arterial blood; the different level between arterial and venous blood indicates the amount of H<sub>2</sub> incorporated and consumed into tissues (Ohsawa et al., 2007).

In a clinical examination, Ono et al. also showed a difference in H<sub>2</sub> concentrations between arterial and venous blood (Ono et al., 2012a).

The gaseous diffusion of H<sub>2</sub> can be monitored inside various tissues by detecting with a specific electrode. For example, H<sub>2</sub> concentration has been monitored within the rat myocardium. The electrode was inserted into the 'at risk' area for infarction to estimate the diffusion of H<sub>2</sub> into the ischemic myocardium area after coronary artery occlusion. H<sub>2</sub> concentration was increased by its diffusion even with coronary artery occlusion (Hayashida et al., 2008).

Moreover, we devised eye drops with dissolved H<sub>2</sub> to directly administer H<sub>2</sub> to the retina, and monitored the time course of changes

in H<sub>2</sub> levels using a needle-shaped hydrogen sensor electrode inserted through the sclera to the vitreous body in rats. H<sub>2</sub> could reach the vitreous body by administering H<sub>2</sub> saturated in normal saline. When H<sub>2</sub> eye drops were administered continuously, approximately 70% H<sub>2</sub> was detected on the ocular surface (Oharazawa et al., 2010).

When water saturated with H<sub>2</sub> was placed into the stomach of a rat, H<sub>2</sub> was detected at several μM in blood (Nagata et al., 2009; Nakashima-Kamimura et al., 2009). Moreover, hepatic H<sub>2</sub> was monitored with a needle-type hydrogen electrode, and hepatic glycogen maintained H<sub>2</sub> after oral administration of H<sub>2</sub>-water, partly explaining why consumption of even a small amount of H<sub>2</sub> over a short dwell time could efficiently improve various disease models (Kamimura et al., 2011).

When seven adult volunteers drank H<sub>2</sub>-water, the H<sub>2</sub> content of their expired breath was measured by gas chromatography with a semiconductor (Shimouchi et al., 2012). The ingestion of H<sub>2</sub>-water rapidly increased breath H<sub>2</sub> content to its maximal level 10 min after ingestion and thereafter decreased to the baseline level within 60 min. The loss of H<sub>2</sub> from the water during the experimental procedures accounted for 3% or less of the H<sub>2</sub>. H<sub>2</sub> release from the skin surface was estimated as approximately 0.1%. Based on the remaining H<sub>2</sub> mass balance, approximately 40% of H<sub>2</sub> that had been drunk was consumed inside the body. This report suggests that exogenous H<sub>2</sub> is at least partially trapped by oxygen radicals, such as •OH (Shimouchi et al., 2012).

### 6. Effects of molecular hydrogen and clinical examinations

#### 6.1. Safety of molecular hydrogen for humans

H<sub>2</sub> has advantages from the aspect of toxicity: H<sub>2</sub> has no cytotoxicity even at high concentration (Abraini et al., 1994; Lillo et al., 1997; Fontanari et al., 2000; Lillo & Parker, 2000). Safety standards have been established for high concentrations of H<sub>2</sub> gas for inhalation since high-pressure H<sub>2</sub> gas is used in deep-diving gas mixes to prevent decompression sickness and arterial gas thrombi (Fontanari et al., 2000). Importantly for clinical use, the safety of H<sub>2</sub> for humans is demonstrated by its application of an extremely high concentration of H<sub>2</sub> gas in Hydrex, an exotic, breathing gas mixture of 49% H<sub>2</sub>, 50% helium and 1% O<sub>2</sub>, which is used to prevent decompression sickness and nitrogen narcosis during very deep technical diving (Abraini et al., 1994; Lillo et al., 1997; Fontanari et al., 2000; Lillo & Parker, 2000). Considering that H<sub>2</sub> is an inert gas and nonfunctional in our body, its lack of toxic effects is easily understandable. As mentioned above, inhalation of 1–4% H<sub>2</sub> gas exhibits great efficacy (Ohsawa et al., 2007; Hayashida et al., 2008).

Many reports have supported that H<sub>2</sub> exhibited great efficacy in extensive disease models regardless of the ingestion methods of H<sub>2</sub> as summarized in Fig. 4. This review article focuses mainly on clinical examinations.

#### 6.2. Protective effects against reperfusion injury

Ischemia/reperfusion induces serious oxidative stress, and its injuries should be considered in many clinical treatments. Inhalation of H<sub>2</sub> gas improved ischemia/reperfusion injuries in cerebral (Ohsawa et al., 2007) and myocardial infarction (Hayashida et al., 2008; Yoshida et al., 2012). Hydrogen-saline protected renal ischemia-reperfusion injury (Wang et al., 2011). All clinical manifestations related to post-cardiac arrest (CA) syndrome are attributed to ischemia/reperfusion injury in various organs, including the brain and heart. H<sub>2</sub> gas inhalation yielded great improvement in survival and the neurological deficit score in post-CA syndrome in a rat model (Hayashida et al., 2012).

H<sub>2</sub> also mitigated damage during the transplantation of various organs by H<sub>2</sub> gas (Buchholz et al., 2008), or H<sub>2</sub>-water (Cardinal et al., 2010) and H<sub>2</sub>-preservation solution (Noda et al., 2013).

Ono et al. intravenously administered H<sub>2</sub>-saline with Edaravone, a clinically approved radical scavenger, in 8 patients with acute brain



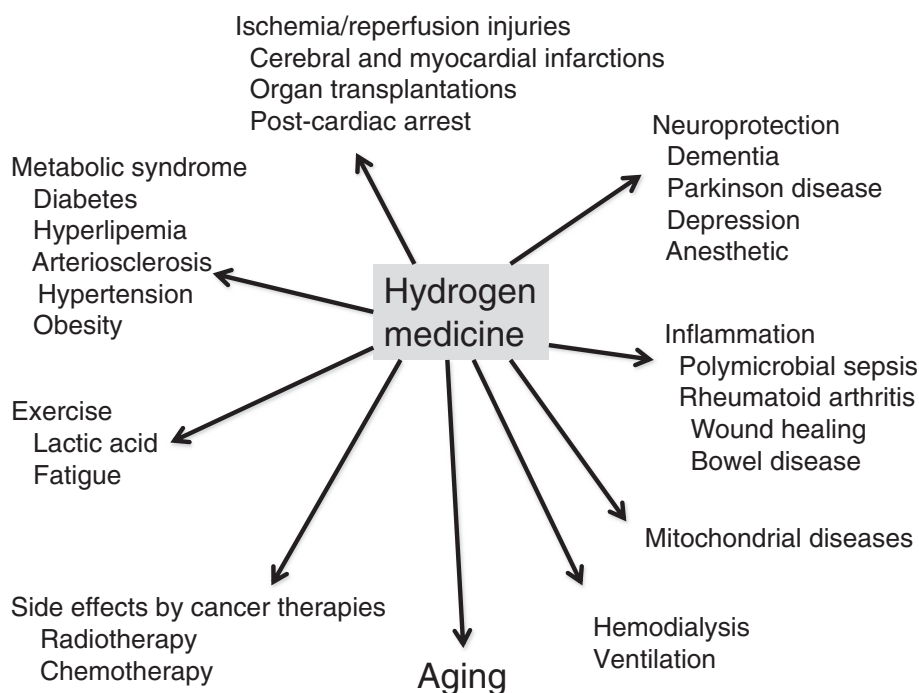


Fig. 4. Summary of potential of various preventive and therapeutic effects of H<sub>2</sub>.

stem infarction and compared magnetic resonance imaging (MRI) indices of 26 patients who received Edaravone alone. The relative diffusion-weighted images (rDWIs), regional apparent diffusion coefficients (rADCs), and pseudo-normalization time of rDWI and rADC were all improved with the combined infusion of H<sub>2</sub> with Edaravone (Ono et al., 2011).

### 6.3. Protective effects against neurodegeneration

Chronic oxidative stress is widely accepted as one of the causes of neurodegeneration, including dementia and Parkinson disease (PD) (Andersen, 2004; Federico et al., 2012). Experimental oxidative stress in the brain can be induced by chronic physical restraint stress and can impair learning and memory (Liu et al., 1996; Abrous et al., 2005). Additionally, neural proliferation in the dentate gyrus of the hippocampus is suppressed by the restraint stress (Abrous et al., 2005). Drinking H<sub>2</sub>-water suppressed the increase in this oxidative stress and prevented this cognitive impairment (Nagata et al., 2009). Moreover, H<sub>2</sub>-water restored neural proliferation in the dentate gyrus of the hippocampus in this model animal (Nagata et al., 2009). Since antidepressants increase adult neurogenesis (Becker & Wojtowicz, 2007; Sahay & Hen, 2007), H<sub>2</sub>-water may be applicable for improving depression and some mental disorders.

In PD, mitochondrial dysfunction and the associated oxidative stress are major causes of dopaminergic cell loss in the substantia nigra (Yoritaka et al., 1996; Schapira, 2008). H<sub>2</sub> in drinking water was given before or after stereotactic surgery for 6-hydroxydopamine-induced nigrostriatal degeneration in a rat model of PD. H<sub>2</sub>-water prevented both the development and progression of nigrostriatal degeneration in rats (Fu et al., 2009). Moreover, drinking H<sub>2</sub>-water also suppressed dopaminergic neuronal loss in another PD mice model induced by MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) (Fujita et al., 2009).

In a placebo-controlled, randomized, double-blind, parallel-group clinical pilot study, the efficacy of H<sub>2</sub>-water in patients with levodopa-medicated PD was assessed (Yoritaka et al., 2013). Participants drank 1 L per day of H<sub>2</sub>-water or pseudo water for 48 weeks. Total Unified Parkinson's Disease Rating Scale (UPDRS) scores in the H<sub>2</sub>-water group (n = 9) improved, whereas UPDRS scores in the placebo group

(n = 8) worsened. Despite the low number of patients and the short duration of the trial, the difference was significant (P < .05).

### 6.4. Preventive effects against metabolic syndrome

Drinking H<sub>2</sub>-water stimulates energy metabolism (Kamimura et al., 2011). H<sub>2</sub>-water significantly alleviated fatty liver in *db/db* mice, which is a type 2 diabetes model mice with obesity, as well as high fat-diet-induced fatty liver in wild-type mice. Long-term H<sub>2</sub>-water drinking significantly decreased fat and body weights, despite no increase in the consumption of diet and water, in *db/db* mice. Moreover, drinking H<sub>2</sub>-water decreased levels of plasma glucose, insulin, and triglyceride by stimulating energy metabolism (Kamimura et al., 2011).

Beneficial roles of H<sub>2</sub>-water in the prevention of potential metabolic syndrome were reported by 3 independent clinical studies as follows.

Kajiyama et al. performed a randomized, double-blinded, placebo-controlled, crossover study in 30 patients with diabetes mellitus type II and 6 patients with impaired glucose tolerance. The patients consumed either 900 mL H<sub>2</sub>-water or placebo water for 8 weeks, with a 12-week washout period. Statistical significance was observed in the improvement of electronegative charge-modified low-density lipoprotein (LDL)-cholesterol, small dense LDL, and urinary 8-isoprostanes. In four of six patients with impaired glucose tolerance, H<sub>2</sub> improved their indexes of the oral glucose tolerance test to a normal level (Kajiyama et al., 2008).

Nakao et al. performed an open-label trial in 20 subjects with potential metabolic syndrome (Nakao et al., 2010b). The participants drank H<sub>2</sub>-water (1.5–2.0 L/day) for 8 weeks and showed an increase in urinary SOD; a decrease in urinary thiobarbituric acid reactive substances (TBARS), a marker of lipid peroxidation; an increase in high-density-lipoprotein (HDL)-cholesterol; and a decrease in total cholesterol.

Song et al. characterized the effects of H<sub>2</sub>-water (0.9–1.0 L/day) on the content, composition, and biological activities of serum lipoproteins in 20 patients with potential metabolic syndrome. Serum analysis showed that drinking H<sub>2</sub>-water for 10 weeks resulted in decreased serum total-cholesterol (TC) and LDL-cholesterol levels. In addition, H<sub>2</sub>-water significantly improved i) protection against LDL oxidation, ii) inhibition of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ )-induced monocyte

adhesion to endothelial cells, iii) stimulation of cholesterol efflux from macrophage foam cells, and iv) protection of endothelial cells from TNF- $\alpha$ -induced apoptosis (Song et al., 2013).

Taken together, these results strongly suggest the benefits of drinking H<sub>2</sub>-water in patients with metabolic syndrome.

### 6.5. Suppressive effects on inflammation

H<sub>2</sub> reduced inflammation in experimental model animals induced by concanavalin A (Kajiya et al., 2009), dextran sodium sulfate (Kajiya et al., 2009), lipopolysaccharide (LPS) (Xu et al., 2012; Chen et al., 2013), Zymosan, an inducer of generalized inflammation (Xie et al., 2010b), and polymicrobial sepsis (G.M. Li et al., 2013). H<sub>2</sub> gas, H<sub>2</sub>-saline and H<sub>2</sub>-water decreased the levels of pro-inflammatory cytokines to suppress inflammation.

Rheumatoid arthritis (RA) is a chronic inflammatory disease characterized by the destruction of bone and cartilage. Ishibashi et al. administered H<sub>2</sub>-water containing 4–5 mg/L H<sub>2</sub> (high H<sub>2</sub>-water) (0.5 L/day) for 4 weeks to 20 patients with RA. Urinary 8-hydroxy-deoxyguanine (8-OHdG) was significantly reduced by 14.3% ( $P < 0.01$ ) on average. Disease activity (DAS28, using C-reactive protein levels also decreased ( $P < 0.01$ ) during the same period. After the washout period, both the urinary 8-OHdG and the mean DAS28 had decreased compared to the end of the drinking period. During the second drinking period, the mean DAS28 was reduced ( $P < 0.01$ ). All the 5 patients with early RA (duration < 12 months) who did not show antibodies against cyclic citrullinated peptides (ACPs) achieved remission, and 4 of them became symptom-free at the end of the study. Thus, the symptoms of RA were significantly improved with H<sub>2</sub>-water (Ishibashi et al., 2012; Ishibashi, 2013).

### 6.6. Mitigation of side effects in treatments of cancers

Inhalation of H<sub>2</sub> gas and drinking H<sub>2</sub>-water improved mortality and body-weight loss caused by treatment with an anti-cancer drug, cisplatin, and alleviated nephrotoxicity in mice. Despite its protective effects against cisplatin-induced toxicity, H<sub>2</sub> did not affect the anti-tumor activity of cisplatin against cancer cell lines in vitro and tumor-bearing mice in vivo (Nakashima-Kamimura et al., 2009).

Kang et al. performed a randomized placebo-controlled clinical study of H<sub>2</sub>-water (0.55–0.65 mM, 1.5–2.0 L/day) for 6 weeks in 49 patients receiving radiation therapy for malignant liver tumors. H<sub>2</sub> suppressed the elevation of total hydroperoxide levels, maintained serum antioxidant capacity, and improved the quality of life (QOL) scores. In particular, H<sub>2</sub>-water efficiently prevented loss of appetite. There was no difference in tumor response to radiotherapy between the two groups (Kang et al., 2011).

### 6.7. Effects on dermatomyositis and mitochondrial diseases

Ito et al. performed an open-label trial of H<sub>2</sub>-water (1.0 L/day) for 12 weeks in 14 patients with muscular diseases, including muscular dystrophies, polymyositis/dermatomyositis, and mitochondrial myopathies. In the open-label trial, significant improvements were observed in the lactate:pyruvate ratio, fasting blood glucose, serum matrix metalloproteinase-3 (MMP3), and triglycerides. In particular, the lactate:pyruvate ratio, which is a sensitive biomarker of a compromised mitochondrial electron transport system, was decreased by 28% in mitochondrial myopathies. In addition, MMP3, which represents the activity of inflammation, was decreased by 27% in dermatomyositis (Ito et al., 2011).

Then, they performed a randomized, double-blind, placebo-controlled, crossover trial of H<sub>2</sub>-water or placebo-dehydrogenized water (0.5 L/day) for 8 weeks in 22 patients with dermatomyositis and mitochondrial myopathies. In the double-blind trial, a statistically significant improvement was observed only in serum lactate in mitochondrial

myopathies by H<sub>2</sub>-water consumption, but the lactate:pyruvate ratio in mitochondrial myopathies and MMP3 in dermatomyositis also tended to be decreased by H<sub>2</sub>-water consumption (Ito et al., 2011).

### 6.8. Mitigating effects of hemodialysis

Nakayama et al. performed an open-label placebo-controlled crossover trial of 12 sessions of hemodialysis in eight patients (Nakayama et al., 2009, 2010) and an open-label trial of 78 sessions of hemodialysis in 21 patients (Nakayama et al., 2010). In both studies, continuous sessions of hemodialysis with H<sub>2</sub>-dialysis solution decreased systolic blood pressure before and after dialysis. In the short-term study, plasma methylguanidine was significantly decreased. In the long-term study, plasma monocyte chemoattractant protein 1 and myeloperoxidase were significantly decreased.

### 6.9. Effects on acute erythematous skin diseases

Ono et al. treated 4 patients with acute erythematous skin diseases with fever and/or pain by intravenous administration of 0.5 L of H<sub>2</sub>-fluid for 30 min for more than 3 days. Erythema of these 4 patients and associated symptoms improved significantly after H<sub>2</sub> treatment. In conclusion, an improvement in acute erythematous skin diseases followed the administration of H<sub>2</sub>-fluid without compromising safety (Ono et al., 2012b).

### 6.10. Effects on exercise

Aoki et al. recruited ten young male soccer players and subjected them to exercise tests and blood sampling. Each subject was examined twice in a crossover double-blind manner; they were given H<sub>2</sub>-water or placebo water for one-week intervals. Subjects were requested to use a cycle ergometer at 75 % maximal oxygen uptake (VO<sub>2</sub>) for 30 min, followed by measurement of peak torque and muscle activity throughout 100 repetitions of maximal isokinetic knee extension. Acute exercise resulted in an increase in blood lactate levels in the subjects given placebo water, whereas oral intake of H<sub>2</sub>-water prevented the elevation of blood lactate during heavy exercise. Peak torque in the placebo water-group significantly decreased during maximal isokinetic knee extension, indicating muscle fatigue (Aoki et al., 2012).

## 7. Possible molecular mechanisms underlying various effects of molecular hydrogen

As mentioned, H<sub>2</sub> has various beneficial effects on animal models and patients; anti-oxidation, anti-inflammation, anti-apoptosis, anti-allergy and stimulation of energy metabolism (Ohta, 2011, 2012). Their mutual relationships are not clear, but the reduction of oxidative stress may primarily lead to various subsequent effects. There remain many unresolved questions regarding the molecular mechanism to fully explain the effects of H<sub>2</sub>. In particular, although H<sub>2</sub> apparently regulates gene expressions and the protein phosphorylation involved in signal transduction, the primary target(s) of H<sub>2</sub> in these regulations has not been identified. Here, possible mechanisms are proposed as summarized in Fig. 5.

### 7.1. Direct reduction of hydroxyl radicals with molecular hydrogen

Considering the reaction rate of •OH with H<sub>2</sub> in dilute aqueous solutions, the reaction rate may be too slow to fully enable a decrease in •OH for exhibiting its beneficial roles (Buxton et al., 1988). Mammalian cells are, however, highly structured with complicated biomembranes and viscous solutions with concentrated multiple components. Since collision frequency is rate-limiting in a viscous environment, the marked diffusion rate of H<sub>2</sub> could be advantageous to overcome the slow reaction rate constant.

H<sub>2</sub> was shown to reduce •OH in an experiment using cultured cells (Ohsawa et al., 2007); however, the reduction of •OH to exhibit therapeutic and preventive effects had not been directly demonstrated at tissue level at this time. Later, it was shown that H<sub>2</sub> eye-drops directly decreased •OH induced by ischemia reperfusion in retinas (Oharazawa et al., 2010). Moreover, it has been demonstrated that, at a tissue level, H<sub>2</sub> neutralized •OH that had been induced by ionizing irradiation in testes, as judged by the decreased HFP signal, and exhibited a radio-protective role of H<sub>2</sub> (Chuai et al., 2012).

•OH is known as a major trigger of the chain reaction of free radicals (Niki, 2009). Once the chain reaction occurs on biomembranes, it continues and expands causing serious damage to cells. H<sub>2</sub> may accumulate in the lipid phase more than in the aqueous phase, especially in unsaturated lipid regions, which are the major target of the initial chain reaction. Thus, H<sub>2</sub> may have an advantage to suppress the chain reaction, which produces lipid peroxide, and leads to the generation of oxidative stress markers, such as 4-hydroxyl-2-nonenal (4-HNE) and malondialdehyde (MDA) (Niki, 2014). Indeed, H<sub>2</sub> decreased these oxidative markers in many studies (Ohsawa et al., 2008; Ning et al., 2013; Zhou et al., 2013). Additionally, •OH can modify deoxy-guanine (dG) to 8-OHdG (Delaney et al., 2012; Kawai et al., 2012). H<sub>2</sub> decreased the level of 8-OHdG in most of the examined patients and animals (Ishibashi et al., 2012; Kawai et al., 2012).

Thus, these observations support that sufficient H<sub>2</sub> can efficiently mitigate tissue oxidation induced by •OH. However, when animals or humans drink H<sub>2</sub>-water, the question remains whether H<sub>2</sub>-water provides a sufficient amount of H<sub>2</sub> to efficiently scavenge •OH, which are continuously generated in normal and disease states.

### 7.2. Direct reduction of peroxynitrite with molecular hydrogen to regulate gene expressions

As another molecular mechanism, scavenging ONOO<sup>-</sup> by H<sub>2</sub> should be considered. ONOO<sup>-</sup> is known to modify tyrosine of proteins to generate nitro-tyrosine (Radi, 2013). Several studies have shown that drinking H<sub>2</sub>-water efficiently decreases nitro-tyrosine in animal models regardless of H<sub>2</sub>-water (Cardinal et al., 2010), H<sub>2</sub> gas (Shinbo et al., 2013) or H<sub>2</sub>-saline (C.H. Chen et al., 2010; Yu et al., 2011; Zhang et al.,

2011; Zhu et al., 2011). Moreover, drinking H<sub>2</sub>-water decreased nitro-tyrosine in patients with RA (Ishibashi et al., 2012). Thus, at least a part of the effect of H<sub>2</sub> can be attributed to the decreased production of nitro-tyrosine of proteins.

Many protein factors involved in transcriptional control are nitrated (-O-NO<sub>2</sub>) or nitrosolated (-S-NO<sub>2</sub>). It is possible that the decrease in -O-NO<sub>2</sub> or -S-NO<sub>2</sub> may regulate various gene expressions (Radi, 2013). However, major targets have not been identified and are under investigation.

### 7.3. Indirect reduction of oxidative stress by regulating gene expression

H<sub>2</sub> reduces oxidative stress not only directly, but indirectly by inducing anti-oxidation systems, including hemeoxygenase-1 (HO-1) (Huang et al., 2010; Park et al., 2010), SOD (Zhai et al., 2013), catalase (Cai et al., 2013) and myeloperoxidase (Zhang et al., 2011). Nrf2 is known to function as a defense system against oxidative stress and various poisons by inducing various genes including HO-1. HO-1, a microsomal enzyme degrading heme to carbon monoxide, free iron, and biliverdin, participates in the cell defense against oxidative stress (Jazwa & Cuadrado, 2010).

In Nrf2-deficient mice, mitigating effects by inhalation of H<sub>2</sub> gas declined in hyperoxic lung injury accompanying with a decrease in HO-1, indicating that H<sub>2</sub> gas can ameliorate hyperoxic lung injury in an Nrf2-dependent manner (Kawamura et al., 2013). Activation of Nrf2 is also required for amelioration of cerebral ischemia-reperfusion injury in rats by H<sub>2</sub> (Zhai et al., 2013).

### 7.4. Regulation of gene expression of pro-inflammatory cytokines and hormones

H<sub>2</sub> seems to display various effects against various pathogenic situations indirectly by up- or down-regulation of gene expression.

In most inflammatory models, H<sub>2</sub> functions as anti-inflammation by decreasing the expressions of pro-inflammatory factors (Ohta, 2011). These pro-inflammatory factors include NF-κB (H. Chen et al., 2010), TNF-α, interleukin (IL)-1β, IL-6, IL-10, IL-12, CCL2 and interferon (INF)-γ, ICAM-1 (Buchholz et al., 2008), PGE2 and PGE2, (Kawasaki

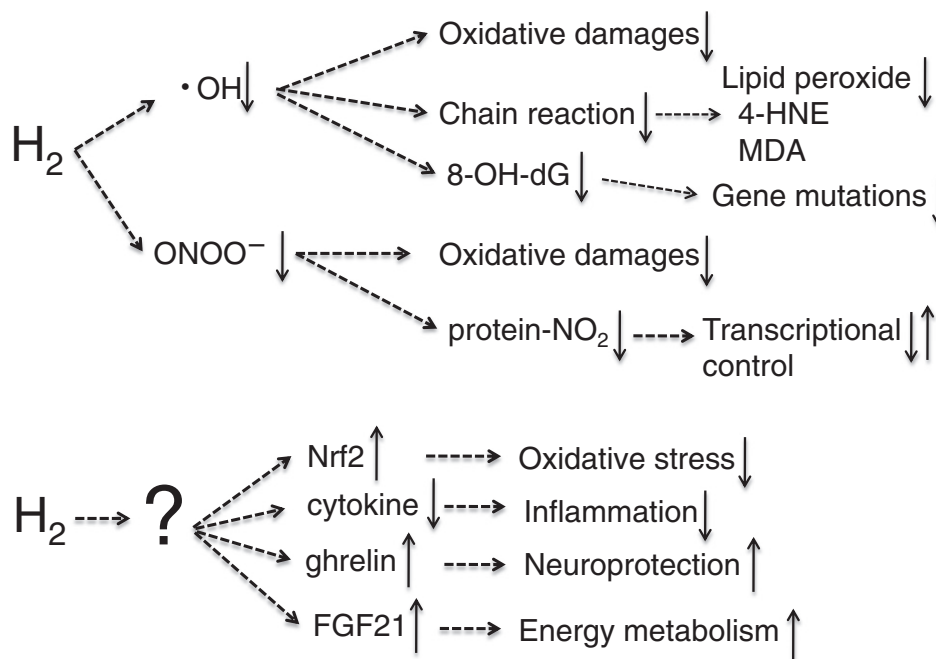


Fig. 5. Possible mechanisms of the marked efficacy by H<sub>2</sub> in various disease models and patients.

et al., 2010), and high mobility group box 1 (HMGB-1) (Xie et al., 2010a),

H<sub>2</sub>-water improves obesity and metabolic parameters as mentioned above. Analysis of gene expression revealed that a hepatic hormone, fibroblast growth factor 21 (FGF21), showed increased expression by drinking H<sub>2</sub>-water. FGF21 functions to stimulate fatty acid and glucose expenditure (Kamimura et al., 2011).

H<sub>2</sub>-water supplementation increased the gastric expression of mRNA encoding ghrelin, a growth hormone secretagogue, and ghrelin secretion. Strikingly, the neuroprotective effect of H<sub>2</sub>-water was abolished by administration of an antagonist to the ghrelin receptor. Thus, a neuroprotective effect of H<sub>2</sub> may be mediated by enhanced production of ghrelin (Matsumoto et al., 2013).

It is unlikely that H<sub>2</sub> could directly react with transcriptional factors to regulate the gene expressions as shown in Fig. 5.

### 7.5. Regulation of other genes

H<sub>2</sub> functions toward the suppression of apoptosis by up-regulation of anti-apoptotic factors, and/or by down-regulation of pro-apoptotic factors against various pathogenic statuses. Indeed, H<sub>2</sub> stimulated expressions of the anti-apoptotic factors of Bcl-2, and Bcl-x<sub>L</sub> (Kawamura et al., 2010) and suppressed the expressions of pro-apoptotic factors including caspase 3 (Cai et al., 2008; Sun et al., 2009), caspase 8 (Kawamura et al., 2010) and caspase 12 (J. Cai et al., 2008). In pro-apoptotic Bax, H<sub>2</sub> did not only down-regulate its gene expression (Kawamura et al., 2010; Huang et al., 2011), but also inhibited the translocation of Bax to mitochondria (Terasaki et al., 2011) by an unknown mechanism.

Expression profiling of rat liver demonstrated that H<sub>2</sub>-water has a minimal effect on the expression levels of individual genes in normal rats (Nakai et al., 2011); however, more apparent up- or down-regulation of individual genes were observed in disease models and patients against the pathogenic stimuli.

H<sub>2</sub> seems to exhibit a variety of phenotypes toward improving many pathogenic states by regulating various gene expressions. Genes up- or down-regulated by H<sub>2</sub> are as follows; MMP2 and MMP9 (C.H. Chen et al., 2010); MMP3 and MMP13 (Hanaoka et al., 2011); brain natriuretic peptide; intercellular-adhesion-molecule-1 (ICAM-1) and myeloperoxidase; cyclooxygenase 2 (COX-2), neuronal nitric oxide synthase (nNOS), and connexins 30 and 43 (Hugyecz et al., 2011); collagen III (Terasaki et al., 2011); and ionized calcium binding adaptor molecule 1 (Iba1) (Sun et al., 2011b).

Additionally, recent reports indicated that ventilation with H<sub>2</sub> significantly increased expression of surfactant-related molecules, ATP synthases and stress-response molecules in lung grafts (Tanaka et al., 2012), and that H<sub>2</sub> reduced mRNA levels of osteoclast-specific markers, including tartrate resistant acid phosphatase, calcitonin receptor, cathepsin K, metalloproteinase-9, carbonic anhydrase type-II, and vacuolar-type H<sup>+</sup>-ATPase (D.Z. Li et al., 2013).

These molecules are, probably, not primary responders to H<sub>2</sub>, but indirectly act to enable the various effects of H<sub>2</sub>. The primary target of H<sub>2</sub> remains unknown.

### 7.6. Modulation of signaling

H<sub>2</sub> attenuates the phosphorylation of FcεR1-associated Lyn and its downstream signaling molecules (Itoh et al., 2009). H<sub>2</sub> inhibits the phosphorylation of ASK1 and its downstream signaling molecules, p38 MAP kinase, JNK, and IκB without affecting ROS production derived from NADPH oxidase. The other study also indicated that H<sub>2</sub> inhibited the phosphorylations of some signal proteins, including MEK, p38, ERK, JNK (Cardinal et al., 2010). Pretreatment with H<sub>2</sub> reduced fatty acid uptake and lipid accumulation after palmitate overload in HepG2 cells, which was associated with inhibition of JNK activation (Iio et al., 2013). H<sub>2</sub> reduced the phosphorylation of p38, extracellular signal-

regulated kinase, JNK, and protein kinases B stimulated with the receptor activator of NFκB ligand-induced osteoclast differentiation (D.M. Li et al., 2013). These studies suggest that H<sub>2</sub> influences some signal transductions as an indirect modulator; however, it is unlikely that H<sub>2</sub> could directly bind to some receptors involved in the signal transductions. The primary target molecule of H<sub>2</sub> has not been identified in these signal transduction pathways.

## 8. Comparison with the other medical gasses

Several medical gasses are expected to provide more effective therapeutic interventions and preventive medicine. Hydrogen sulfide (H<sub>2</sub>S), carbon monoxide (CO), and •NO are extremely toxic molecules; however, they play important roles as signaling molecules (Kimura, 2010; Motterlini & Otterbein, 2010). CO is poisonous even in low concentrations due to its ability to interfere with oxygen delivery, whereas endogenous CO is important in the physiological functioning of organs and possesses anti-inflammatory properties (Motterlini & Otterbein, 2010). In contrast, H<sub>2</sub> is advantageous to have no cytotoxicity.

In the past decade, there has been marked, rapid growth in the knowledge of gaseous molecules. Gas inhalation as disease therapy has received recent interest (Szabó, 2007; Kajimura et al., 2010). It has recently been revealed that heme-based proteins play central roles in their generation- and reception-mechanisms as the primary target of these gas molecules. An integrated approach to the interaction of these gasses revealed the physiological significance of H<sub>2</sub>S, CO, and •NO on mitochondrial cytochrome c oxidase, a key target and central mediator of mitochondrial respiration (Kajimura et al., 2010). As far as briefly examined (Ohsawa et al., 2007), H<sub>2</sub> did not reduce the oxidized heme of cytochrome c. Thus, the primary target of H<sub>2</sub> seems to differ from that of the other medical gasses.

Regarding the interaction between H<sub>2</sub> and the other toxic medical gasses, combined therapy with H<sub>2</sub> and CO demonstrated enhanced therapeutic efficacy via both anti-oxidant and anti-inflammatory mechanisms, and may be a clinically feasible approach for preventing ischemia/reperfusion injury in the myocardium (Nakao et al., 2010a).

Breathing NO plus H<sub>2</sub> during ischemia/reperfusion reduced the infarct size and maintained cardiac function, and reduced the generation of myocardial nitrotyrosine associated with •NO inhalation. Administration of •NO plus H<sub>2</sub> gasses for inhalation may be useful for planned coronary interventions or for the treatment of ischemia/reperfusion injury (Shinbo et al., 2013).

The production of •NO, H<sub>2</sub>S or CO is catalyzed by •NO synthases, cystathionine γ-lyase and cystathionine β-synthase or HO-1, respectively (Kashfi & Olson, 2013). In contrast, mammalian cells have no enzyme for producing intracellular H<sub>2</sub>.

## 9. Concluding remarks

This article reviewed the progress of hydrogen medicine from its initiation toward clinical applications. H<sub>2</sub> is easily applicable because it has no adverse effects and great efficacy on nearly all pathogenic states involved in oxidative stress and inflammation. Indeed, the clinical effects of H<sub>2</sub> were positive in patients with more than 10 various diseases. Since most pharmacological drugs specifically act on their targets, H<sub>2</sub> seems to differ from conventional drugs because of its extensive and various effects. H<sub>2</sub> has great potential for preventive and therapeutic applications in many diseases owing to its great efficacy and its novel concept.

## Conflict of Interest Statement

The author is a Chief Scientific Officer of MitoGene, LLC (Little Rock, AR, USA).



## References

- Abrajain, J. H., Gardette-Chauffour, M. C., Martinez, E., Rostain, J. C., & Lemaire, C. (1994). Psychophysiological reactions in humans during an open sea dive to 500 m with a hydrogen-helium-oxygen mixture. *J Appl Physiol* 76, 1113–1118.
- Abrous, D. N., Koehl, M., & Le Moal, M. (2005). Adult neurogenesis: from precursors to network and physiology. *Physiol Rev* 85, 523–569.
- Agarwal, R. (2005). Smoking, oxidative stress and inflammation: impact on resting energy expenditure in diabetic nephropathy. *BMC Nephrol* 6, 13.
- Andersen, J. K. (2004). Oxidative stress in neurodegeneration: cause or consequence? *Nat Med* 10, S18–S25 (Suppl.).
- Aoki, K., Nakao, A., Adachi, T., Matsui, Y., & Miyakawa, S. (2012). Pilot study: Effects of drinking hydrogen-rich water on muscle fatigue caused by acute exercise in elite athletes. *Med Gas Res* 2, 12.
- Bagul, P. K., & Banerjee, S. K. (2013). Insulin resistance, oxidative stress and cardiovascular complications: role of sirtuins. *Curr Pharm Des* 19, 5663–5677.
- Bashan, N., Kovsan, J., Kachko, I., Ovadia, H., & Rudich, A. (2009). Positive and negative regulation of insulin signaling by reactive oxygen and nitrogen species. *Physiol Rev* 89, 27–71.
- Becker, S., & Wojtowicz, J. M. (2007). A model of hippocampal neurogenesis in memory and mood disorders. *Trends Cogn Sci* 11, 70–76.
- Bell, E. L., Klimova, T. A., Eisenbart, J., Moraes, C. T., Murphy, M. P., Budinger, G. R., et al. (2007). The Qo site of the mitochondrial complex III is required for the transduction of hypoxic signaling via reactive oxygen species production. *J Cell Biol* 177, 1029–1036.
- Bell, E. L., Klimova, T. A., Eisenbart, J., Schumacker, P. T., & Chandel, N. S. (2007). Mitochondrial reactive oxygen species trigger hypoxia-inducible factor-dependent extension of the replicative life span during hypoxia. *Mol Cell Biol* 27, 5737–5745.
- Bjelakovic, G., Nikolova, D., Gluud, L. L., Simonetti, R. G., & Gluud, C. (2007). Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *JAMA* 297, 842–857.
- Brambilla, D., Mancuso, C., Scuderi, M. R., Bosco, P., Cantarella, G., Lempereur, L., et al. (2008). The role of antioxidant supplement in immune system, neoplastic, and neurodegenerative disorders: a point of view for an assessment of the risk/benefit profile. *Nutr J* 7, 29.
- Brewer, A. C., Mustafi, S. B., Murray, T. V., Rajasekaran, N. S., & Benjamin, I. J. (2013). Reductive stress linked to small HSPs, G6PD, and Nrf2 pathways in heart disease. *Antioxid Redox Signal* 18, 1114–1127.
- Buchholz, B. M., Kaczorowski, D. J., Sugimoto, R., Yang, R., Wang, Y., Billiar, T. R., et al. (2008). Hydrogen inhalation ameliorates oxidative stress in transplantation induced intestinal graft injury. *Am J Transplant* 8, 2015–2024.
- Buxton, G. V., Greenstock, C. L., Helman, W. P., & Ross, A. B. (1988). Critical view of rate constants for reactions of hydrated electrons, hydrogen atoms and hydroxyl radicals ( $\cdot\text{OH}/\cdot\text{OH}^-$ ) in aqueous solution. *J Phys Chem Ref Data* 17, 513–886.
- Cai, J., Kang, Z., Liu, K., Liu, W., Li, R., Zhang, J. H., et al. (2009). Neuroprotective effects of hydrogen saline in neonatal hypoxia-ischemia rat model. *Brain Res* 1256, 129–137.
- Cai, J., Kang, Z., Liu, W. W., Luo, X., Qiang, S., Zhang, J. H., et al. (2008). Hydrogen therapy reduces apoptosis in neonatal hypoxia-ischemia rat model. *Neurosci Lett* 441, 167–172.
- Cai, W. W., Zhang, M. H., Yu, Y. S., & Cai, J. H. (2013). Treatment with hydrogen molecule alleviates TNF $\alpha$ -induced cell injury in osteoblast. *Mol Cell Biochem* 373, 1–9.
- Cardinal, J. S., Zhan, J., Wang, Y., Sugimoto, R., Tsung, A., McCurry, K. R., et al. (2010). Oral hydrogen water prevents chronic allograft nephropathy in rats. *Kidney Int* 77, 101–109.
- Chandel, N. S., Trzyna, W. C., McClintock, D. S., & Schumacker, P. T. (2000). Role of oxidants in NF- $\kappa$ B activation and TNF- $\alpha$  gene transcription induced by hypoxia and endotoxin. *J Immunol* 165, 1013–1021.
- Chandel, N. S., Vander Heiden, M. G., Thompson, C. B., & Schumacker, P. T. (2000). Redox regulation of p53 during hypoxia. *Oncogene* 19, 3840–3848.
- Chen, C. H., Manaenko, A., Zhan, Y., Liu, W. W., Ostrowski, R. P., Tang, J., et al. (2010). Hydrogen gas reduced acute hyperglycemia-enhanced hemorrhagic transformation in a focal ischemia rat model. *Neuroscience* 169, 402–414.
- Chen, H., Sun, Y. P., Li, Y., Liu, W. W., Xiang, H. G., Fan, L. Y., et al. (2010). Hydrogen-rich saline ameliorates the severity of L-arginine-induced acute pancreatitis in rats. *Biochem Biophys Res Commun* 393, 308–313.
- Chen, H. G., Xie, K. L., Han, H. Z., Wang, W. N., Liu, D. Q., Wang, G. L., et al. (2013). Heme oxygenase-1 mediates the anti-inflammatory effect of molecular hydrogen in LPS-stimulated RAW 264.7 macrophages. *Int J Surg* 11, 1060–1066.
- Chuai, Y., Gao, F., Li, B., Zhao, L., Qian, L., Cao, F., et al. (2012). Hydrogen-rich saline attenuates radiation-induced male germ cell loss in mice through reducing hydroxyl radicals. *Biochem J* 442, 49–56.
- Collins, Y., Chouchani, E. T., James, A. M., Menger, K. E., Cocheme, H. M., & Murphy, M. P. (2012). Mitochondrial redox signalling at a glance. *J Cell Sci* 125, 801–806.
- Delaney, S., Jarem, D. A., Volle, C. B., & Yennie, C. J. (2012). Chemical and biological consequences of oxidatively damaged guanine in DNA. *Free Radic Res* 46, 420–441.
- Droge, W. (2002). Free radicals in the physiological control of cell function. *Physiol Rev* 82, 47–95.
- El Assar, M., Angulo, J., & Rodriguez-Manas, L. (2013). Oxidative stress and vascular inflammation in aging. *Free Radic Biol Med* 65, 380–401.
- Endo, J., Sano, M., Katayama, T., Hishiki, T., Shinmura, K., Morizane, S., et al. (2009). Metabolic remodeling induced by mitochondrial aldehyde stress stimulates tolerance to oxidative stress in the heart. *Circ Res* 105, 1118–1127.
- Federico, A., Cardaioli, E., Da Pozzo, P., Formichi, P., Gallus, G. N., & Radi, E. (2012). Mitochondria, oxidative stress and neurodegeneration. *J Neurol Sci* 322, 254–262.
- Ferrari, R., Ceconi, C., Curello, S., Cargnoni, A., Pasini, E., & Visioli, O. (1991). The occurrence of oxidative stress during reperfusion in experimental animals and men. *Cardiovasc Drugs Ther* 5(Suppl. 2), 277–287.
- Finkel, T. (1998). Oxygen radicals and signaling. *Curr Opin Cell Biol* 10, 248–253.
- Finkel, T., & Holbrook, N. J. (2000). Oxidants, oxidative stress and the biology of ageing. *Nature* 408, 239–247.
- Fontanari, P., Badier, M., Guillot, C., Tomei, C., Burnet, H., Gardette, B., et al. (2000). Changes in maximal performance of inspiratory and skeletal muscles during and after the 7.1-MPa Hydra 10 record human dive. *Eur J Appl Physiol* 81, 325–328.
- Fritsch, J., Lenz, O., & Friedrich, B. (2013). Structure, function and biosynthesis of O(2)-tolerant hydrogenases. *Nat Rev Microbiol* 11, 106–114.
- Fu, Y., Ito, M., Fujita, Y., Ichihara, M., Masuda, A., Suzuki, Y., et al. (2009). Molecular hydrogen is protective against 6-hydroxydopamine-induced nigrostriatal degeneration in a rat model of Parkinson's disease. *Neurosci Lett* 453, 81–85.
- Fujita, K., Seike, T., Yutsudo, N., Ohno, M., Yamada, H., Yamaguchi, H., et al. (2009). Hydrogen in drinking water reduces dopaminergic neuronal loss in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *PLoS One* 4, e7247.
- Gan, L., & Johnson, J. A. (2013). Oxidative damage and the Nrf2-ARE pathway in neurodegenerative diseases. *Biochim Biophys Acta* (Epub ahead of print).
- Garg, A. D., Dudek, A. M., Ferreira, G. B., Verfaillie, T., Vandenabeele, P., Krysko, D. V., et al. (2013). ROS-induced autophagy in cancer cells assists in evasion from determinants of immunogenic cell death. *Autophagy* 9, 1292–1307.
- Grassi, D., Desideri, G., Ferri, L., Aggio, A., Tiberti, S., & Ferri, C. (2010). Oxidative stress and endothelial dysfunction: say no to cigarette smoking! *Curr Pharm Des* 16, 2539–2550.
- Hackam, D. G. (2007). Review: antioxidant supplements for primary and secondary prevention do not decrease mortality. *ACP J Club* 147, 4.
- Halliwell, B., & Gutteridge, J. M. (1992). Biologically relevant metal ion-dependent hydroxyl radical generation. An update. *FEBS Lett* 307, 108–112.
- Hanaoka, T., Kamimura, N., Yokota, T., Takai, S., & Ohta, S. (2011). Molecular hydrogen protects chondrocytes from oxidative stress and indirectly alters gene expressions through reducing peroxynitrite derived from nitric oxide. *Med Gas Res* 1, 18.
- Harma, M. I., Harma, M., & Erel, O. (2006). Measuring plasma oxidative stress biomarkers in sport medicine. *Eur J Appl Physiol* 97, 505 (author reply 506–508).
- Hayashida, K., Sano, M., Kamimura, N., Yokota, T., Suzuki, M., Maekawa, Y., et al. (2012). H<sub>2</sub> gas improves functional outcome after cardiac arrest to an extent comparable to therapeutic hypothermia in a rat model. *J Am Heart Assoc* 1, e003459.
- Hayashida, K., Sano, M., Ohsawa, I., Shinmura, K., Tamaki, K., Kimura, K., et al. (2008). Inhalation of hydrogen gas reduces infarct size in the rat model of myocardial ischemia-reperfusion injury. *Biochem Biophys Res Commun* 373, 30–35.
- Hercberg, S., Kesse-Guyot, E., Druenes-Pecollo, N., Touvier, M., Favier, A., Latino-Martel, P., et al. (2010). Incidence of cancers, ischemic cardiovascular diseases and mortality during 5-year follow-up after stopping antioxidant vitamins and minerals supplements: a postintervention follow-up in the SU.VI.MAX Study. *Int J Cancer* 127, 1875–1881.
- Huang, C. S., Kawamura, T., Toyoda, Y., & Nakao, A. (2010). Recent advances in hydrogen research as a therapeutic medical gas. *Free Radic Res* 44, 971–982.
- Huang, C. S., Kawamura, T., Peng, X., Tochigi, N., Shigemura, N., Billiar, T. R., et al. (2011). Hydrogen inhalation reduced epithelial apoptosis in ventilator-induced lung injury via a mechanism involving nuclear factor- $\kappa$ B activation. *Biochem Biophys Res Commun* 408, 253–258.
- Hugyecz, M., Mracsók, E., Hertelendy, P., Farkas, E., Domoki, F., & Bari, F. (2011). Hydrogen supplemented air inhalation reduces changes of prooxidant enzyme and gap junction protein levels after transient global cerebral ischemia in the rat hippocampus. *Brain Res* 1404, 31–38.
- Iio, A., Ito, M., Itoh, T., Terazawa, R., Fujita, Y., Nozawa, Y., et al. (2013). Molecular hydrogen attenuates fatty acid uptake and lipid accumulation through downregulating CD36 expression in HepG2 cells. *Med Gas Res* 3, 6.
- Ishibashi, T. (2013). Molecular hydrogen: new antioxidant and anti-inflammatory therapy for rheumatoid arthritis and related diseases. *Curr Pharm Des* 19, 6375–6381.
- Ishibashi, T., Sato, B., Rikitake, M., Seo, T., Kurokawa, R., Hara, Y., et al. (2012). Consumption of water containing a high concentration of molecular hydrogen reduces oxidative stress and disease activity in patients with rheumatoid arthritis: an open-label pilot study. *Med Gas Res* 2, 27.
- Ito, M., Ibi, T., Sahashi, K., Ichihara, M., & Ohno, K. (2011). Open-label trial and randomized, double-blind, placebo-controlled, crossover trial of hydrogen-enriched water for mitochondrial and inflammatory myopathies. *Med Gas Res* 1, 24.
- Itoh, T., Fujita, Y., Ito, M., Masuda, A., Ohno, K., Ichihara, M., et al. (2009). Molecular hydrogen suppresses Fc $\epsilon$ RI-mediated signal transduction and prevents degranulation of mast cells. *Biochem Biophys Res Commun* 389, 651–656.
- Jazwa, A., & Cuadrado, A. (2010). Targeting heme oxygenase-1 for neuroprotection and neuroinflammation in neurodegenerative diseases. *Curr Drug Targets* 11, 1517–1531.
- Kajimura, M., Fukuda, R., Bateman, R. M., Yamamoto, T., & Suematsu, M. (2010). Interactions of multiple gas-transducing systems: hallmarks and uncertainties of CO, NO, and H<sub>2</sub>S gas biology. *Antioxid Redox Signal* 13, 157–192.
- Kajiyama, M., Sato, K., Silva, M. J., Ouhara, K., Do, P. M., Shanmugam, K. T., et al. (2009). Hydrogen from intestinal bacteria is protective for Concanavalin A-induced hepatitis. *Biochem Biophys Res Commun* 386, 316–321.
- Kajiyama, M., Silva, M. J., Sato, K., Ouhara, K., & Kawai, T. (2009). Hydrogen mediates suppression of colon inflammation induced by dextran sodium sulfate. *Biochem Biophys Res Commun* 386, 11–15.
- Kajiyama, S., Hasegawa, G., Asano, M., Hosoda, H., Fukui, M., Nakamura, N., et al. (2008). Supplementation of hydrogen-rich water improves lipid and glucose metabolism in patients with type 2 diabetes or impaired glucose tolerance. *Nutr Res* 28, 137–143.
- Kamimura, N., Nishimaki, K., Ohsawa, I., & Ohta, S. (2011). Molecular Hydrogen Improves Obesity and Diabetes by Inducing Hepatic FGF21 and Stimulating Energy Metabolism in db/db Mice. *Obesity (Silver Spring)* 19, 1396–1403.

- Kang, K. M., Kang, Y. N., Choi, I. B., Gu, Y., Kawamura, T., Toyoda, Y., et al. (2011). Effects of drinking hydrogen-rich water on the quality of life of patients treated with radiotherapy for liver tumors. *Med Gas Res* 1, 11.
- Kashfi, K., & Olson, K. R. (2013). Biology and therapeutic potential of hydrogen sulfide and hydrogen sulfide-releasing chimeras. *Biochem Pharmacol* 85, 689–703.
- Kawai, D., Takaki, A., Nakatsuka, A., Wada, J., Tamaki, N., Yasunaka, T., et al. (2012). Hydrogen-rich water prevents progression of nonalcoholic steatohepatitis and accompanying hepatocarcinogenesis in mice. *Hepatology* 56, 912–921.
- Kawamura, T., Huang, C. S., Tochigi, N., Lee, S., Shigemura, N., Billiar, T. R., et al. (2010). Inhaled hydrogen gas therapy for prevention of lung transplant-induced ischemia/reperfusion injury in rats. *Transplantation* 90, 1344–1351.
- Kawamura, T., Wakabayashi, N., Shigemura, N., Huang, C. S., Masutani, K., Tanaka, Y., et al. (2013). Hydrogen gas reduces hyperoxic lung injury via the Nrf2 pathway in vivo. *Am J Physiol Lung Cell Mol Physiol* 304, L646–L656.
- Kawasaki, H., Guan, J., & Tamama, K. (2010). Hydrogen gas treatment prolongs replicative lifespan of bone marrow multipotential stromal cells in vitro while preserving differentiation and paracrine potentials. *Biochem Biophys Res Commun* 397, 608–613.
- Kim, Y. W., & Byzova, T. V. (2014). Oxidative stress in angiogenesis and vascular disease. *Blood* 123, 625–631.
- Kimura, H. (2010). Hydrogen sulfide: from brain to gut. *Antioxid Redox Signal* 12, 1111–1123.
- Kubota, M., Shimmura, S., Kubota, S., Miyashita, H., Kato, N., Noda, K., et al. (2011). Hydrogen and N-acetyl-L-cysteine rescue oxidative stress-induced angiogenesis in a mouse corneal alkali-burn model. *Invest Ophthalmol Vis Sci* 52, 427–433.
- Li, L., Ishdorj, G., & Gibson, S. B. (2012). Reactive oxygen species regulation of autophagy in cancer: implications for cancer treatment. *Free Radic Biol Med* 53, 1399–1410.
- Li, G. M., Ji, M. H., Sun, X. J., Zeng, Q. T., Tian, M., Fan, Y. X., et al. (2013). Effects of hydrogen-rich saline treatment on polymicrobial sepsis. *J Surg Res* 181, 279–286.
- Li, D. Z., Zhang, Q. X., Dong, X. X., Li, H. D., & Ma, X. (2013). Treatment with hydrogen molecules prevents RANKL-induced osteoclast differentiation associated with inhibition of ROS formation and inactivation of MAPK, AKT and NF- $\kappa$ B pathways in murine RAW264.7 cells. *J Bone Miner Metab* (Epub ahead of print).
- Lillo, R. S., & Parker, E. C. (2000). Mixed-gas model for predicting decompression sickness in rats. *J Appl Physiol* 89, 2107–2116.
- Lillo, R. S., Parker, E. C., & Porter, W. R. (1997). Decompression comparison of helium and hydrogen in rats. *J Appl Physiol* 82, 892–901.
- Lin, M. T., & Beal, M. F. (2006). Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 443, 787–795.
- Lipinski, B. (2011). Hydroxyl radical and its scavengers in health and disease. *Oxid Med Cell Longev* 2011, 809696.
- Liu, H., Colavitti, R., Rovira, I. I., & Finkel, T. (2005). Redox-dependent transcriptional regulation. *Circ Res* 97, 967–974.
- Liu, J., Wang, X., Shigenaga, M. K., Yeo, H. C., Mori, A., & Ames, B. N. (1996). Immobilization stress causes oxidative damage to lipid, protein, and DNA in the brain of rats. *FASEB J* 10, 1532–1538.
- Mates, J. M., Segura, J. A., Alonso, F. J., & Marquez, J. (2012). Oxidative stress in apoptosis and cancer: an update. *Arch Toxicol* 86, 1649–1665.
- Matsumoto, A., Yamafuji, M., Tachibana, T., Nakabeppu, Y., Noda, M., & Nakaya, H. (2013). Oral 'hydrogen water' induces neuroprotective ghrelin secretion in mice. *Sci Rep* 3, 3273.
- Motterlini, R., & Otterbein, L. E. (2010). The therapeutic potential of carbon monoxide. *Nat Rev Drug Discov* 9, 728–743.
- Nagata, K., Nakashima-Kamimura, N., Mikami, T., Ohsawa, I., & Ohta, S. (2009). Consumption of molecular hydrogen prevents the stress-induced impairments in hippocampus-dependent learning tasks during chronic physical restraint in mice. *Neuropsychopharmacology* 34, 501–508.
- Nagatani, K., Nawashiro, H., Takeuchi, S., Tomura, S., Otani, N., Osada, H., et al. (2013). Safety of intravenous administration of hydrogen-enriched fluid in patients with acute cerebral ischemia: initial clinical studies. *Med Gas Res* 3, 13.
- Nakai, Y., Sato, B., Ushima, S., Okada, S., Abe, K., & Arai, S. (2011). Hepatic oxidoreduction-related genes are upregulated by administration of hydrogen-saturated drinking water. *Biosci Biotechnol Biochem* 75, 774–776.
- Nakao, A., Kaczorowski, D. J., Wang, Y., Cardinal, J. S., Buchholz, B. M., Sugimoto, R., et al. (2010a). Amelioration of rat cardiac cold ischemia/reperfusion injury with inhaled hydrogen or carbon monoxide, or both. *J Heart Lung Transplant* 29, 544–553.
- Nakao, A., Toyoda, Y., Sharma, P., Evans, M., & Guthrie, N. (2010b). Effectiveness of hydrogen rich water on antioxidant status of subjects with potential metabolic syndrome—an open label pilot study. *J Clin Biochem Nutr* 46, 140–149.
- Nakashima-Kamimura, N., Mori, T., Ohsawa, I., Asoh, S., & Ohta, S. (2009). Molecular hydrogen alleviates nephrotoxicity induced by an anti-cancer drug cisplatin without compromising anti-tumor activity in mice. *Cancer Chemother Pharmacol* 64, 753–761.
- Nakayama, M., Kabayama, S., Nakano, H., Zhu, W. J., Terawaki, H., Nakayama, K., et al. (2009). Biological effects of electrolyzed water in hemodialysis. *Nephron Clin Pract* 112, c9–c15.
- Nakayama, M., Nakano, H., Hamada, H., Itami, N., Nakazawa, R., & Ito, S. (2010). A novel bioactive haemodialysis system using dissolved dihydrogen (H<sub>2</sub>) produced by water electrolysis: a clinical trial. *Nephrol Dial Transplant* 25, 3026–3033.
- Niki, E. (2009). Lipid peroxidation: physiological levels and dual biological effects. *Free Radic Biol Med* 47, 469–484.
- Niki, E. (2014). Biomarkers of lipid peroxidation in clinical material. *Biochim Biophys Acta* 1840, 809–817.
- Ning, Y., Shang, Y., Huang, H., Zhang, J., Dong, Y., Xu, W., et al. (2013). Attenuation of cigarette smoke-induced airway mucus production by hydrogen-rich saline in rats. *PLoS One* 8, e83429.
- Noda, K., Shigemura, N., Tanaka, Y., Kawamura, T., Hyun Lim, S., Kokubo, K., et al. (2013). A novel method of preserving cardiac grafts using a hydrogen-rich water bath. *J Heart Lung Transplant* 32, 241–250.
- Oharazawa, H., Igarashi, T., Yokota, T., Fujii, H., Suzuki, H., Machide, M., et al. (2010). Protection of the retina by rapid diffusion of hydrogen: administration of hydrogen-loaded eye drops in retinal ischemia-reperfusion injury. *Invest Ophthalmol Vis Sci* 51, 487–492.
- Ohsawa, I., Ishikawa, M., Takahashi, K., Watanabe, M., Nishimaki, K., Yamagata, K., et al. (2007). Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. *Nat Med* 13, 688–694.
- Ohsawa, I., Nishimaki, K., Yamagata, K., Ishikawa, M., & Ohta, S. (2008). Consumption of hydrogen water prevents atherosclerosis in apolipoprotein E knockout mice. *Biochem Biophys Res Commun* 377, 1195–1198.
- Ohta, S. (2011). Recent progress toward hydrogen medicine: potential of molecular hydrogen for preventive and therapeutic applications. *Curr Pharm Des* 17, 2241–2252.
- Ohta, S. (2012). Molecular hydrogen is a novel antioxidant to efficiently reduce oxidative stress with potential for the improvement of mitochondrial diseases. *Biochim Biophys Acta* 1820, 586–594.
- Ono, H., Nishijima, Y., Adachi, N., Sakamoto, M., Kudo, Y., Kaneko, K., et al. (2012a). A basic study on molecular hydrogen (H<sub>2</sub>) inhalation in acute cerebral ischemia patients for safety check with physiological parameters and measurement of blood H<sub>2</sub> level. *Med Gas Res* 2, 21.
- Ono, H., Nishijima, Y., Adachi, N., Sakamoto, M., Kudo, Y., Nakazawa, J., et al. (2012b). Hydrogen(H<sub>2</sub>) treatment for acute erythematous skin diseases. A report of 4 patients with safety data and a non-controlled feasibility study with H<sub>2</sub> concentration measurement on two volunteers. *Med Gas Res* 2, 14.
- Ono, H., Nishijima, Y., Adachi, N., Tachibana, S., Chitoku, S., Mukaiharu, S., et al. (2011). Improved brain MRI indices in the acute brain stem infarct sites treated with hydroxyl radical scavengers, Edaravone and hydrogen, as compared to Edaravone alone. A non-controlled study. *Med Gas Res* 1, 12.
- Park, D. J., Agarwal, A., & George, J. F. (2010). Heme oxygenase-1 expression in murine dendritic cell subpopulations: effect on CD8+ dendritic cell differentiation in vivo. *Am J Pathol* 176, 2831–2839.
- Radi, R. (2013). Peroxynitrite, a stealthy biological oxidant. *J Biol Chem* 288, 26464–26472.
- Reuter, S., Gupta, S. C., Chaturvedi, M. M., & Aggarwal, B. B. (2010). Oxidative stress, inflammation, and cancer: how are they linked? *Free Radic Biol Med* 49, 1603–1616.
- Ristow, M., & Zarse, K. (2010). How increased oxidative stress promotes longevity and metabolic health: The concept of mitochondrial hormesis (mitohormesis). *Exp Gerontol* 45, 410–418.
- Sahay, A., & Hen, R. (2007). Adult hippocampal neurogenesis in depression. *Nat Neurosci* 10, 1110–1115.
- Salganik, R. I. (2001). The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients and the human population. *J Am Coll Nutr* 20, 464S–472S (discussion 473S–475S).
- Sauer, H., Wartenberg, M., & Hescheler, J. (2001). Reactive oxygen species as intracellular messengers during cell growth and differentiation. *Cell Physiol Biochem* 11, 173–186.
- Schapiro, A. H. (2008). Mitochondria in the aetiology and pathogenesis of Parkinson's disease. *Lancet Neurol* 7, 97–109.
- Schoenfeld, M. P., Ansari, R. R., Nakao, A., & Wink, D. (2012). A hypothesis on biological protection from space radiation through the use of new therapeutic gases as medical countermeasures. *Med Gas Res* 2, 8.
- Schoenfeld, M. P., Ansari, R. R., Zakrajsek, J. F., Billiar, T. R., Toyoda, Y., Wink, D. A., et al. (2011). Hydrogen therapy may reduce the risks related to radiation-induced oxidative stress in space flight. *Med Hypotheses* 76, 117–118.
- Setsukinai, K., Urano, Y., Kakinuma, K., Majima, H. J., & Nagano, T. (2003). Development of novel fluorescence probes that can reliably detect reactive oxygen species and distinguish specific species. *J Biol Chem* 278, 3170–3175.
- Shimouchi, A., Nose, K., Shirai, M., & Kondo, T. (2012). Estimation of molecular hydrogen consumption in the human whole body after the ingestion of hydrogen-rich water. *Adv Exp Med Biol* 737, 245–250.
- Shinbo, T., Kokubo, K., Sato, Y., Hagiri, S., Hataishi, R., Hirose, M., et al. (2013). Breathing nitric oxide plus hydrogen gas reduces ischemia-reperfusion injury and nitrotyrosine production in murine heart. *Am J Physiol Heart Circ Physiol* 305, H542–H550.
- Song, G., Li, M., Sang, H., Zhang, L., Li, X., Yao, S., et al. (2013). Hydrogen-rich water decreases serum LDL-cholesterol levels and improves HDL function in patients with potential metabolic syndrome. *J Lipid Res* 54, 1884–1893.
- Steinhubl, S. R. (2008). Why have antioxidants failed in clinical trials? *Am J Cardiol* 101, 14D–19D.
- Sun, Q., Cai, J., Liu, S., Liu, Y., Xu, W., Tao, H., et al. (2011a). Hydrogen-rich saline provides protection against hyperoxic lung injury. *J Surg Res* 165, e43–e49.
- Sun, Q., Cai, J., Zhou, J., Tao, H., Zhang, J. H., Zhang, W., et al. (2011b). Hydrogen-rich saline reduces delayed neurologic sequelae in experimental carbon monoxide toxicity. *Crit Care Med* 39, 765–769.
- Sun, Q., Kang, Z., Cai, J., Liu, W., Liu, Y., Zhang, J. H., et al. (2009). Hydrogen-rich saline protects myocardium against ischemia/reperfusion injury in rats. *Exp Biol Med* (Maywood) 234, 1212–1219.
- Szabó, C. (2007). Hydrogen sulphide and its therapeutic potential. *Nat Rev Drug Discov* 6, 917–935.
- Tanaka, Y., Shigemura, N., Kawamura, T., Noda, K., Isse, K., Stolz, D. B., et al. (2012). Profiling molecular changes induced by hydrogen treatment of lung allografts prior to procurement. *Biochem Biophys Res Commun* 425, 873–879.
- Tanriverdi, H., Evrengul, H., Kuru, O., Tanriverdi, S., Selegi, D., Enli, Y., et al. (2006). Cigarette smoking induced oxidative stress may impair endothelial function and coronary blood flow in angiographically normal coronary arteries. *Circ J* 70, 593–599.
- Terasaki, Y., Ohsawa, I., Terasaki, M., Takahashi, M., Kunugi, S., Dedong, K., et al. (2011). Hydrogen therapy attenuates irradiation-induced lung damage by reducing oxidative stress. *Am J Physiol Lung Cell Mol Physiol* 301, L415–L426.

- Tormos, K. V., Anso, E., Hamanaka, R. B., Eisenbart, J., Joseph, J., Kalyanaraman, B., et al. (2011). Mitochondrial complex III ROS regulate adipocyte differentiation. *Cell Metab* 14, 537–544.
- Tsakagoshi, H., Busch, W., & Benfey, P. N. (2010). Transcriptional regulation of ROS controls transition from proliferation to differentiation in the root. *Cell* 143, 606–616.
- Turrens, J. F. (2003). Mitochondrial formation of reactive oxygen species. *J Physiol* 552, 335–344.
- Vaziri, N. D., & Rodriguez-Iturbe, B. (2006). Mechanisms of disease: oxidative stress and inflammation in the pathogenesis of hypertension. *Nat Clin Pract Nephrol* 2, 582–593.
- Wang, F., Yu, G., Liu, S. Y., Li, J. B., Wang, J. F., Bo, L. L., et al. (2011). Hydrogen-rich saline protects against renal ischemia/reperfusion injury in rats. *J Surg Res* 167, e339–e344.
- West, A. P., Brodsky, I. E., Rahner, C., Woo, D. K., Erdjument-Bromage, H., Tempst, P., et al. (2011). TLR signalling augments macrophage bactericidal activity through mitochondrial ROS. *Nature* 472, 476–480.
- Westerblad, H., & Allen, D. G. (2011). Emerging roles of ROS/RNS in muscle function and fatigue. *Antioxid Redox Signal* 15, 2487–2499.
- Xie, K., Yu, Y., Pei, Y., Hou, L., Chen, S., Xiong, L., et al. (2010). Protective effects of hydrogen gas on murine polymicrobial sepsis via reducing oxidative stress and HMGB1 release. *Shock* 34, 90–97.
- Xie, K., Yu, Y., Zhang, Z., Liu, W., Pei, Y., Xiong, L., et al. (2010). Hydrogen gas improves survival rate and organ damage in zymosan-induced generalized inflammation model. *Shock* 34, 495–501.
- Xu, Z., Zhou, J., Cai, J., Zhu, Z., Sun, X., & Jiang, C. (2012). Anti-inflammation effects of hydrogen saline in LPS activated macrophages and carrageenan induced paw oedema. *J Inflamm (Lond)* 9, 2.
- Yoritaka, A., Hattori, N., Uchida, K., Tanaka, M., Stadtman, E. R., & Mizuno, Y. (1996). Immunohistochemical detection of 4-hydroxynonenal protein adducts in Parkinson disease. *Proc Natl Acad Sci U S A* 93, 2696–2701.
- Yoritaka, A., Takashi, M., Hirayama, M., Nakahara, T., Ohta, S., & Hattori, N. (2013). Pilot study of H<sub>2</sub> therapy in Parkinson's disease: a randomized double-blind placebo-controlled trial. *Mov Disord* 28, 836–839.
- Yoshida, A., Asanuma, H., Sasaki, H., Sanada, S., Yamazaki, S., Asano, Y., et al. (2012). H<sub>2</sub> mediates cardioprotection via involvements of K(ATP) channels and permeability transition pores of mitochondria in dogs. *Cardiovasc Drugs Ther* 26, 217–226.
- Yu, P., Wang, Z., Sun, X., Chen, X., Zeng, S., Chen, L., et al. (2011). Hydrogen-rich medium protects human skin fibroblasts from high glucose or mannitol induced oxidative damage. *Biochem Biophys Res Commun* 409, 350–355.
- Zhai, X., Chen, X., Shi, J., Shi, D., Ye, Z., Liu, W., et al. (2013). Lactulose ameliorates cerebral ischemia-reperfusion injury in rats by inducing hydrogen by activating Nrf2 expression. *Free Radic Biol Med* 65, 731–741.
- Zhang, Y., Sun, Q., He, B., Xiao, J., Wang, Z., & Sun, X. (2011). Anti-inflammatory effect of hydrogen-rich saline in a rat model of regional myocardial ischemia and reperfusion. *Int J Cardiol* 148, 91–95.
- Zhou, H., Fu, Z., Wei, Y., Liu, J., Cui, X., Yang, W., et al. (2013). Hydrogen inhalation decreases lung graft injury in brain-dead donor rats. *J Heart Lung Transplant* 32, 251–258.
- Zhou, R., Yazdi, A. S., Menu, P., & Tschopp, J. (2011). A role for mitochondria in NLRP3 inflammasome activation. *Nature* 469, 221–225.
- Zhu, W. J., Nakayama, M., Mori, T., Nakayama, K., Katoh, J., Murata, Y., et al. (2011). Intake of water with high levels of dissolved hydrogen (H<sub>2</sub>) suppresses ischemia-induced cardio-renal injury in Dahl salt-sensitive rats. *Nephrol Dial Transplant* 26, 2112–2118.